

PARSING THE PROSENCEPHALON

Murielle Rallu, Joshua G. Corbin and Gord Fishell

The forebrain, or prosencephalon, consists of the diencephalon and the telencephalon. The diencephalon is the conduit for ascending sensory information, whereas the telencephalon is the highest-order processor of neural function, and is consequently the most complex region of the nervous system. In this review, we discuss how fate restrictions, starting from the induction of neural character, result in the sequential specification of anterior neural tissue, forebrain and telencephalon, and finally dorsoventral patterning. Rather than relying on novel signalling pathways, the complexity of the mature brain seems to result from the unique ordering of signals used widely during development.

HOMEODOMAIN

A 60-amino-acid DNA-binding domain that comprises three α -helices and is found in many transcription factors.

BASIC HELIX-LOOP-HELIX

(bHLH). A structural motif present in many transcription factors that is characterized by two α -helices separated by a loop. The helices mediate dimerization, and the adjacent basic region is required for DNA binding.

NODE

A major organizing centre in primitive-streak-stage embryos that regulates pattern formation. It is known as Hensen's node in chick and the Spemann organizer in frog.

Developmental Genetics Program and the Department of Cell Biology, The Skirball Institute of Biomolecular Medicine, New York University Medical Center, 540 First Avenue, New York, New York 10016, USA. Correspondence to G.F. e-mail: fishell@saturn.med.nyu.edu doi:10.1038/nrn989

Over the past decade, our understanding of regional patterning in the vertebrate telencephalon, and of the mechanisms that mediate its assembly, has moved forward significantly. This progress can be attributed largely to insights gained from studies in *Drosophila*^{1,2}, and to extrapolation from mechanisms that are known to pattern other levels of the neuraxis³. In general, the lesson has been that transient signalling centres produce diffusible cues that create positional information. Recipient cells translate these signals through the induction of combinatorial codes of transcription factors, and as a result, they acquire specific cellular identities⁴.

The clearest example of a factor that provides positional information within the telencephalon is sonic hedgehog (**Shh**)⁵. Specifically, gain- and loss-of-function analyses of Shh signalling in the telencephalon have indicated that, as at other levels of the neuraxis, this diffusible signalling molecule is essential for the expression of characteristic ventral identities, as well as the repression of complementary dorsal fates⁶⁻⁹. A cadre of specific genes that bestow cellular identity in the telencephalon in response to Shh signalling has been found¹⁰. For the most part, these genes have been identified as the vertebrate homologues of those that establish neuronal populations in *Drosophila*². So far, the proteins encoded by these genes have fallen primarily into two broad classes of transcription factors — those containing a characteristic DNA-binding HOMEODOMAIN sequence and those that contain a BASIC HELIX-LOOP-HELIX (bHLH) motif. Despite the similarities in the underlying logic of the mechanisms that establish different neural regions in

vertebrates, or the brain in invertebrates, it is becoming clear that the telencephalon cannot simply be considered as an anterior-localized spinal cord or an elaborate fly head. In this review, we summarize what has been discovered so far about the extrinsic and intrinsic programmes that participate in the patterning of the telencephalon, and how these programmes interact. We argue that progress in our understanding of the telencephalon has entered a new phase, in which the key insights will come from direct examination of the vertebrate telencephalon, rather than from indirect inferences based on findings in other systems.

Induction of anterior neural character

The early patterning of both anterior and posterior neural tissues is mediated through signals that emanate from the primitive NODE OR ORGANIZER. Studies in mammals indicate that, in addition to the organizer, the anterior visceral endoderm (AVE) is required for head induction and maintenance¹¹⁻¹⁵ (FIG. 1). The AVE is the extra-embryonic tissue that underlies the future neural plate OR EPIBLAST¹⁵ (FIG. 1). Removal of the AVE from mouse embryos at early stages of gastrulation leads to a loss or reduction of forebrain marker expression¹⁵. Also, several mutants that lack genes that are normally expressed in the AVE (for example, *Hex1*, *Lim1* and *Otx2*) fail to develop anterior structures, including the forebrain¹⁶. Finally, transplantation of the mouse AVE into chick embryos results in the expression of forebrain markers in the epiblast. However, it is unclear whether the AVE has an active or a passive role in establishing

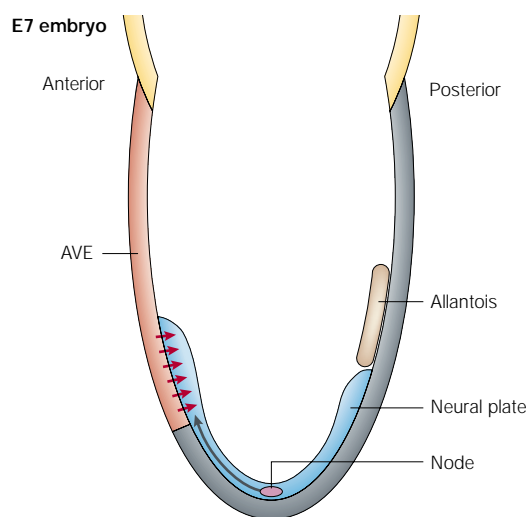


Figure 1 | Signals and tissues involved in inducing anterior neural character. Schematic of a mouse embryo at the early headfold stage. Signals that come from the node establish gross anterior pattern (black arrow). The anterior visceral endoderm (AVE), together with the node, acts to induce and/or maintain anterior neural character. The AVE is located beneath the future neural plate and expresses molecules, such as *cerberus* and *dickkopf* (red arrows), that inhibit factors that would otherwise act to posteriorize the anterior neural plate. This figure represents the end stage at which these signals are acting. E7, embryonic day 7.

anterior neural structures¹². Transplantation of the chick hypoblast beneath the lateral epiblast/ectoderm (the equivalent of the AVE) can transiently induce anterior markers, but cannot sustain them. Furthermore, its removal does not prevent the expression of forebrain markers¹⁷, indicating that the signals for head induction or maintenance might be present in different structures in mammals and birds.

Although the source of forebrain inducers might vary between different species, numerous lines of evidence indicate that the signalling mechanisms that specify the anterior neural tissue are widely conserved. Considerable molecular and genetic data support a model proposed by Nieuwkoop¹⁸, who suggested that nascent neural tissue adopts an anterior identity by default (a process that he referred to as activation), and that the posterior nervous system is subsequently generated through a process that he called 'transformation'. Recently, this model has been revisited in the light of a wealth of new evidence¹². Studies in fish and frogs have implicated *Wnt*, retinoids, and fibroblast growth factor (FGF) as 'posteriorizing' factors^{19–22}, indicating that inhibitors of these diffusible signals are responsible for maintaining anterior neural identity. With respect to the antagonism of *Wnt* signalling, two proteins with this activity, *cerberus* and *dickkopf*, are expressed in the anterior endoderm of frogs (the equivalent of the AVE) (FIG. 2), and can induce the formation of a head (but not a trunk) when misexpressed in frog embryos^{23,24} (see note added in proof). The transforming growth factor- β (TGF- β)-related family of proteins, such as

bone morphogenetic proteins (BMPs), activins and *Nodal*, also seem to play a part in this process. For example, mutants in which *Nodal* signalling is compromised have an enlarged telencephalon, indicating that the blockade of *Nodal* signalling is involved in forebrain development²⁵. Interestingly, *cerberus* and *dickkopf*, which, in addition to their ability to block *Wnt* signalling, also function as *Nodal* or BMP antagonists^{23,24}, are only two of the many proteins that block these signalling pathways. These include *chordin*, *noggin*, *follistatin* and the Frizzled-related protein *Frzb*^{26–28}, all of which seem to act in the specification of the forebrain. Indeed, in mice that lack both *chordin* and *noggin* gene activities, the expression of AVE markers is induced correctly but is not maintained, and anterior neural fates are ultimately lost.

To complicate matters, it is clear that each of these proteins has many functions in addition to their roles in anteroposterior (AP) patterning of the nervous system; FGF is a potent mesoderm inducer, BMPs have a role in dorsoventral (DV) patterning of the early embryo, and both *Wnt* and *Nodal* signalling act in the establishment of the AP axis before neurulation^{12,14,16,19,21,25,26,28}. Whether or not the role of these proteins in establishing anterior versus posterior neural structures can be neatly divided from their other functions is unclear at present.

Specification of telencephalic character
Subsequent to anterior neural induction, the cells at the junction between the anterior neural and non-neural ectoderm — the anterior neural ridge (ANR) or anterior neural boundary (ANB) — have an important role in promoting telencephalic development within the forebrain territory. Both in mice²⁹ and in fish³⁰, ablation of these cells prevents the expression of telencephalic markers such as *Bf1* and *Emx1*. In zebrafish, transplantation of these cells into the midbrain can induce telencephalic and suppress midbrain marker expression in a cell-non-autonomous fashion. Other studies in zebrafish have indicated that the inhibition of *Wnt* signalling is a crucial step in the specification of the telencephalon and in the subdivision of the forebrain into telencephalic, optic and diencephalic territories. The *masterblind* mutation, which inactivates the *Axin* gene (a negative regulator of *Wnt* signalling)³¹, transforms the telencephalon into diencephalic tissue³². Furthermore, Houart *et al.*³³ have shown that the telencephalon-inducing activity of the ANB cells is mediated by *Tlc*, a novel secreted Frizzled-related protein that acts as an extracellular antagonist of *Wnt* signalling (FIG. 2). *Tlc* not only mimics the activity of ANB cells, but *tlc* gene function is also required for telencephalic development in zebrafish³³. Therefore, similar to the induction of the anterior neural tissues, the specification of telencephalic identity might require the inhibition of posteriorizing (diencephalic) signals, including, but probably not limited to, *Wnt* signalling.

Dorsoventral regionalization of the telencephalon
At the headfold stage (E8.0, four to eight somites), the mouse telencephalic anlage lies within the anterior third of the paired, downward-folded leaves of the neural plate, and the two sides of the anlage meet at the anterior

EPIBLAST
The outer layer of a blastula, which gives rise to the ectoderm after gastrulation.

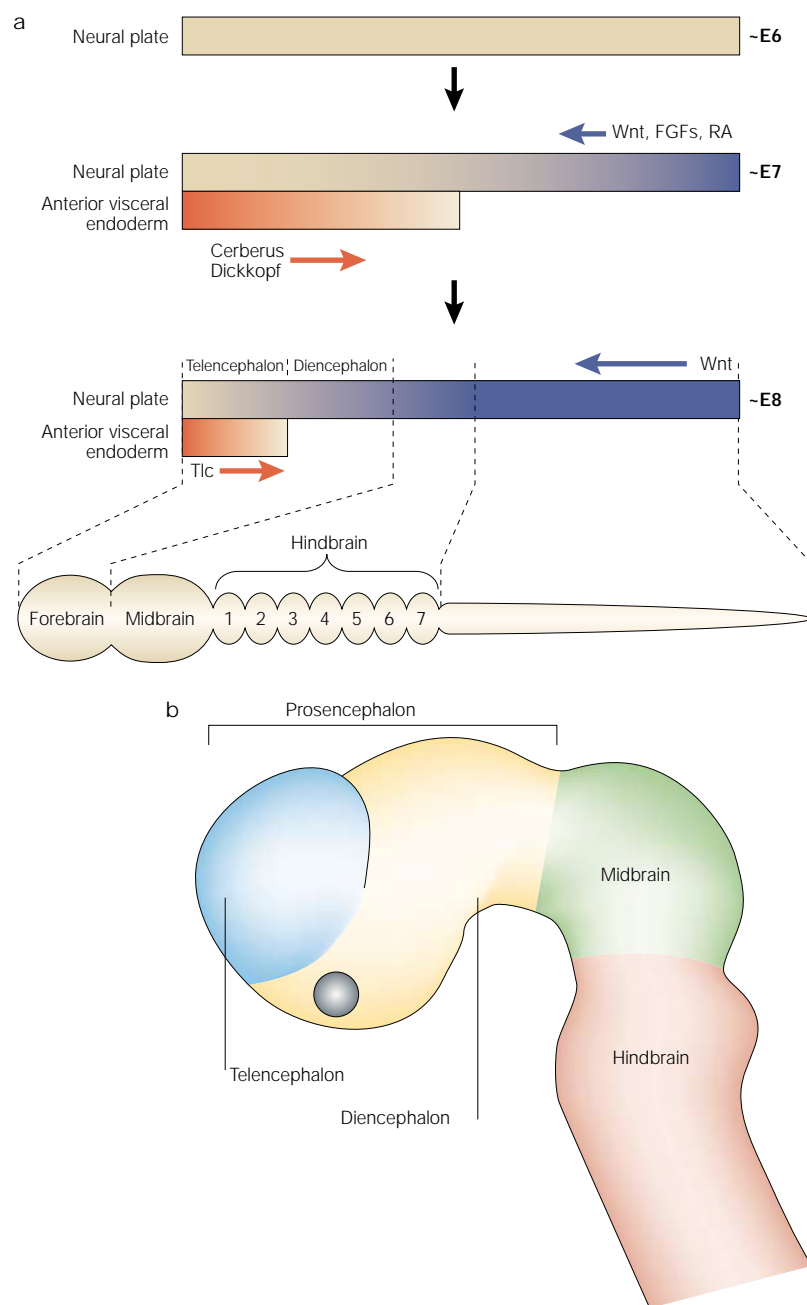


Figure 2 | Progressive specification of the telencephalon. a | Neural induction results in the formation of the neural plate. Markers expressed throughout the early neural plate will ultimately become restricted to anterior domains of the central nervous system. A number of molecules, including the Wnts, fibroblast growth factors (FGFs) and retinoids (RA), can function at this stage of development to induce posterior character in the neural plate. Conversely, antagonists of these factors, including cerberus and dickkopf, are expressed in the anterior visceral endoderm and act to maintain and stabilize the anterior neural plate character. Subsequently, the anterior neural domain is subdivided as a result of graded Wnt signalling. A key aspect of this process appears to be the expression of the Wnt antagonist Tlc, a Frizzled-related protein. E, embryonic day. **b** | Side view of the brain of a mouse embryo at around embryonic day 10 (E10), showing the main subdivisions.

midline. By coupling a dramatic set of morphogenetic movements with extensive proliferation, the telencephalon is transformed, by around E9 (the 20-somite stage), into a set of paired vesicles, complete with regionally restricted DV markers. These regional markers

presage the morphological appearance of discrete dorsal (cortex), lateral (lateral ganglionic eminence or LGE) and ventral (medial ganglionic eminence or MGE) proliferative zones, which appear two days later, around embryonic day 11 (E11). When, and how, these telencephalic subdivisions are specified remain open questions. By contrast, DV patterning within the more posterior regions of the neural tube has been explained in considerable detail.

Shh signalling. Until recently, ideas about how the DV pattern is established in the forebrain have been largely borrowed from insights gained in the spinal cord. In both regions, a central player in this process is Shh. However, recent data have indicated that, despite some obvious similarities, the mechanisms that are used to establish DV patterning in the telencephalon differ from those used at more posterior regions of the neuraxis.

Shh signalling is crucial for ventral patterning at all levels of the nervous system^{34,35}. Loss- and gain-of-function analyses in several species have shown that the Shh protein is necessary and sufficient for the development of ventral neural structures and the expression of associated neural markers. Embryos that lack Shh fail to form normal ventral telencephalic structures, and they show markedly reduced expression of ventral markers^{6,9,36}. Furthermore, ectopic expression of Shh is sufficient to induce ventral telencephalic marker expression, both *in vitro* and *in vivo*⁷⁻⁹. However, precisely when Shh is required for ventral telencephalic patterning, and whether it acts to differentially specify distinct ventral telencephalic cell fates, is poorly understood. Furthermore, although Shh is expressed by a variety of anterior tissues throughout development, the precise source of Shh activity for telencephalic patterning has not been clearly identified.

The earliest site of Shh expression appears at around E7.5 in the midline mesoderm of the head process³⁷. Shortly afterwards, both Shh and Indian hedgehog (Ihh) messenger RNA appear in the primitive node. As neurulation progresses, both the PRECHORDAL PLATE and the anterior MESENDOERM come to express Shh. Finally, just before the onset of neurogenesis, the expression of Shh occurs in the hypothalamus, and then in the ventral telencephalon itself³⁸. Extirpation and genetic mutations that affect specific subdomains of Shh expression have been shown to perturb ventral telencephalic organization, but they have not yet allowed a clear determination of which sources of Shh are required for which aspects of ventral telencephalic patterning. By contrast, *in vitro* antibody perturbation experiments have given hints as to the temporal requirements for Shh during telencephalic development. These experiments indicate that the ventral-most telencephalic character (MGE-like fate) is acquired in response to Shh signals during gastrula stages³⁹, whereas later exposure of telencephalic tissue seems to be involved in the acquisition of ventrolateral telencephalic fate⁷. Gain-of-function studies have produced similar results^{7,39}.

PRECHORDAL PLATE

A tissue derived from the node, which lies at the rostral tip of the notochord.

MESENODERM

Embryonic tissue that gives rise both to mesoderm and endoderm.

An important difference between the spinal cord and the telencephalon was revealed by *in vitro* experiments in which explants were exposed to various concentrations of Shh. Whereas spinal cord explants show concentration-dependent changes in the ventral genes that are induced by Shh, the fates induced by Shh in the telencephalon depend on the timing, rather than on the concentration, of Shh exposure^{7,39}. The observation that the competence of the telencephalon to respond to Shh changes during development indicates that further mechanisms are involved in the establishment of DV patterning in this area. In accordance with this idea, careful analysis of ventral telencephalic markers in *Shh*-null mutants has raised the possibility that another pathway acts in parallel with Shh to specify DV pattern within the telencephalon. Specifically, the lateral telencephalic fates (LGE) seem to be correctly specified, although they are shifted ventrally at the expense of the most ventral (MGE) fates.

Function of Gli genes. To understand how Shh affects telencephalic patterning, the role of the three mammalian Gli genes, *Gli1*, *Gli2* and *Gli3*, must be considered. Gli genes are homologous to the *Drosophila* gene *cubitus interruptus (ci)*, and genetic analysis has shown that all Hedgehog signalling is mediated through *ci* in flies, and similarly by the Gli genes in vertebrates^{40,41}. *Gli1* and *Gli2* act principally as activators, whereas *Gli3* acts mainly as a repressor⁴¹. With respect to the telencephalon, compound-mutant mice that lack *Gli1* and *Gli2* gene function develop relatively normally⁴². By contrast, in *Gli3* mutants, ventral telencephalic markers expand dorsally into the cortex^{9,43,44}. The fact that opposite telencephalic phenotypes are observed in *Shh* and *Gli3* mutants indicates that the balance between *Shh* and *Gli3* gene function is crucial in the establishment of DV patterning within the telencephalon (FIG. 3).

To explore this hypothesis, we have recently analysed telencephalic patterning in *Shh/Gli3* compound mutants⁹ (FIG. 3). Consistent with work in the spinal cord⁴⁵ and limb^{46,47}, we found that Shh is likely to act through the inhibition of Gli3 repressor activity. Indeed, the removal of one or both alleles of *Gli3* is sufficient to restore the ventral telencephalic gene expression that is lost in *Shh* mutants. Notably, in the absence of both *Shh* and *Gli3* gene function, not only are the lateral fates specified correctly (as in the spinal cord), but so is both the level and localization of *Nkx2.1* gene expression, consistent with the restoration of the MGE, the most ventral part of the telencephalon.

The hedgehog pathway in vertebrates is mediated through multiple ligands and Gli proteins, but all hedgehog signalling requires the function of the transmembrane protein *Smo*, a homologue of *Drosophila Smoothed*. Mice that lack both *Gli3* and *Smo* gene functions produce a similar phenotype to that observed in *Shh/Gli3* compound mutants. Together, these results indicate that Shh is the only hedgehog ligand that functions in the DV patterning of the telencephalon.

One caveat to these findings is that relatively few markers of DV pattern have been identified in the telencephalon, compared with the spinal cord. It remains possible that further analysis with more specific telencephalic markers will reveal that the rescue of DV pattern in these compound mutants is incomplete. For example, the possibility that a subdomain of *Nkx2.1* expression (equivalent to the floorplate in the spinal cord) remains absent in the double mutants cannot be ruled out. This cautionary note aside, these results indicate that the role of *Shh* in DV patterning varies at different levels of the neuraxis. Specifically, *Gli2* is required in the spinal cord for the specification of V3 interneurons and floorplate. By contrast, the loss of *Gli2* has no effect on patterning in the telencephalon. Given that

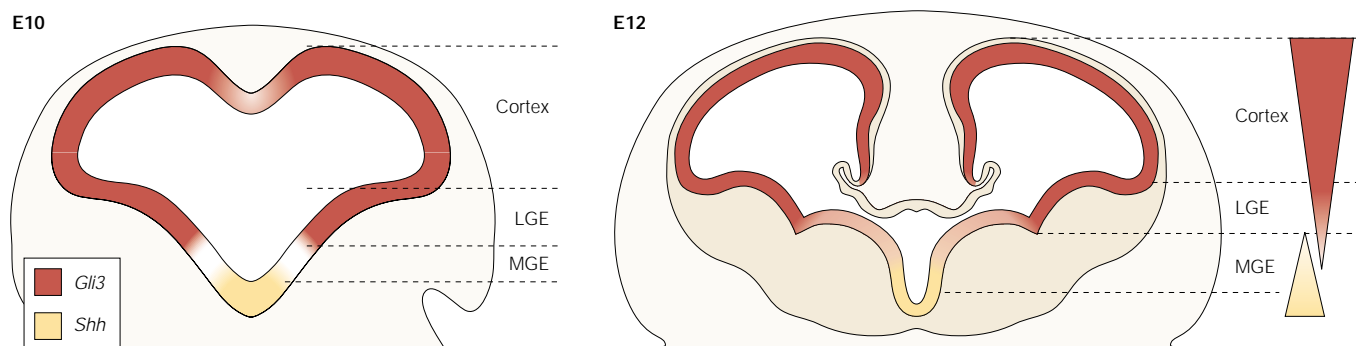


Figure 3 | Model of genetic interactions between Shh and Gli3 in patterning the mouse telencephalon. Schematic representation of a coronal section through an embryonic day 10 (E10; on the left) and E12 (on the right) mouse telencephalon, highlighting different domains along the dorsoventral (DV) axis. The expression domains of *Gli3* (in red) and sonic hedgehog (*Shh*; in yellow) are shown at early and late stages of telencephalic development. These genes maintain a complementary pattern of expression throughout development, and genetic analysis shows that their activities strongly antagonize one another. Specifically, in the absence of *Shh* gene function, the telencephalon is strongly ventralized, whereas in the absence of *Gli3* gene function, the telencephalon is strongly dorsalized. In the absence of both *Gli3* and *Shh*, the general aspects of DV patterning are rescued. The notable exceptions are the dorsal midline structures, which are lost in all three mutant genotypes: *Shh*^{-/-}, *Gli3*^{-/-} and *Shh*^{-/-}*Gli3*^{-/-}. These data indicate the existence of an unknown hedgehog-independent pathway that acts in parallel with *Shh* in the establishment of telencephalic DV pattern. MGE, medial ganglionic eminence; LGE, lateral ganglionic eminence.

several studies argue that Gli3 functions primarily as a repressor^{45–47}, whereas Gli1 and Gli2 act either primarily or wholly as activators⁴⁸, this indicates that only the repressive functions of Shh signalling are required for telencephalic patterning.

These results show that *Shh* is dispensable for the general DV patterning of the telencephalon, provided that *Gli3* function is also abolished. This strongly implies that other signalling pathways act in parallel with *Shh* to regionalize the telencephalon. Although the identities of the Shh-independent signals are unknown, there are several obvious candidates on the basis of patterning mechanisms detected in other studies, primarily in the spinal cord.

Other signalling pathways. BMP signalling, in addition to its role in the specification of dorsal neural tube cell types^{49–51}, also seems to influence the patterning of ventral cell types. Application of BMP proteins to spinal cord explants alters the response of neural progenitors to the Shh protein and results in a dorsal shift in their cellular identity. Moreover, the addition of BMP inhibitors in the same assay seems to potentiate the response to Shh signalling. Therefore, it seems that, in the spinal cord, Shh and BMP signalling pathways converge and potentially limit the actions of each other along the DV axis. Similarly, BMPs are expressed in and around the dorsal telencephalon, and they seem to have a role in the dorsal patterning of this tissue^{43,52–56}. Furthermore, Shh and BMP signalling have been shown to cooperate to induce ventral diencephalic identity⁵⁷. So, early **BMP7** expression in the prechordal plate, or **BMP4** in the PRESOMITIC MESODERM, might also be responsible for Shh-independent patterning. Perhaps the best evidence for a requirement for BMPs in telencephalic patterning comes from an analysis of the zebrafish *swirl* mutant, which is a *Bmp2b* null. The dorsal telencephalic gene *Emx1* is lost in *swirl* mutants, although it is unclear whether this indicates a loss of dorsal structures or loss of the telencephalon as a whole⁵⁸.

In addition to BMPs, Wnts are also involved in dorsal telencephalic development; in particular, in the specification of the hippocampus^{55,59,60}. Recent work also indicates that inhibition of the Wnt pathway is an important feature in the formation of the telencephalon (see above). So, in addition to specifying the telencephalon as a whole, graded inhibition of Wnt signalling might also act to establish DV identity within the telencephalon.

In the spinal cord, a retinoid-activated pathway has been implicated in the generation of the ventrolateral V0 and V1 interneurons, independently of the Shh pathway⁶¹. In the telencephalon, markers of retinoid synthesis and signalling are expressed in the LGE and the developing striatum^{62,63}. Furthermore, retinoid signalling has been shown to regulate striatal neuronal differentiation. However, earlier expression of the retinoids (in the lateral cranial mesoderm⁶⁴) is more likely to have a role in the initial patterning of the telencephalon.

Other candidates that have been implicated in telencephalic patterning are the FGFs. Studies of zebrafish and mice have implicated **FGF3** and **FGF8** signalling in establishing regional patterning in the SUBPALLIAL region of the telencephalon^{29,58,65}. Finally, experiments in zebrafish indicate that Shh and Nodal signalling might interact⁶⁶. Indeed, *Shh* overexpression is able to restore ventral gene expression in the telencephalon of several zebrafish mutants with deficiencies in Nodal signalling. So, it seems that Nodal can mediate forebrain patterning through the regulation of the hedgehog signalling pathway. It is therefore likely that many or all of these signalling pathways act in some capacity in the DV patterning of the telencephalon, but their precise roles, and whether these pathways are acting in concert with or in parallel to Shh signalling, will require further examination.

Downstream patterning mechanisms

In response to the reception of Shh and other extrinsic signals, cells express specific transcription factors. These, in turn, activate intrinsic cellular programmes, which cause progenitors to adopt specific cell fates. Our understanding of the downstream effectors of cell identity remains, at best, rudimentary, but it is known that many of the genes that act in response to extrinsic cues belong to a set of homeodomain-class transcription factors that are conserved across divergent species. For example, the homeodomain proteins that act to pattern the *Drosophila* nerve cord are homologous to those that act to delineate regional cell types in the vertebrate spinal cord and telencephalon^{67,68}. In *Drosophila*, these genes include *vnd* (*ventral nervous system defective*), *ind* (*intermediate neuroblasts defective*) and *msh* (*muscle segment homeobox*), whereas patterning of the vertebrate spinal cord is dependent on the ventral expression of *Nkx2.2* and *Nkx6.1*, and on the intermediate expression of *Dbx1* and *Dbx2* (REF. 69), which are homologues of the *Drosophila H2.0* gene⁷⁰. Similarly, future telencephalic territories can be defined early in the development of this region by the expression of a distinct set of homeodomain genes: *Nkx2.1*, *Gsh1*, *Gsh2* and *Pax6*, which are homologues of the *Drosophila* genes *vnd*, *ind* and *eyeless* (*ey*), respectively^{71–74}. These mouse genes provide some of the earliest markers of dorsal (*Pax6*), intermediate (*Gsh2*) and ventral (*Nkx2.1*) domains of the telencephalon¹⁰ (J.G.C. and G.F., unpublished observations; FIG. 4). Furthermore, these genes are essential for the normal development of the regions in which they are expressed. So, across divergent species, although the extrinsic signalling mechanisms that establish positional cues vary, the result of such signalling manifests itself in the expression of a conserved set of regionally expressed transcription factors.

Of the proteins that act to pattern the vertebrate and invertebrate nervous system, the *Nkx/vnd* family of genes is exceptionally well conserved, in terms of both expression and function⁶⁷. In the absence of the *Nkx/vnd* genes, the fates of the ventral-most cells in the spinal cord⁷⁵ and the *Drosophila* nerve cord are transformed to that of their nearest dorsal neighbours^{71,72}.

PRESOMITIC MESODERM
Precursor unsegmented mesoderm, which generates somites on segmentation.

SUBPALLIAL
Belonging to the base of the telencephalon. The subpallium consists primarily of the basal ganglia, including the striatum, globus pallidus, and parts of the septum and amygdala.

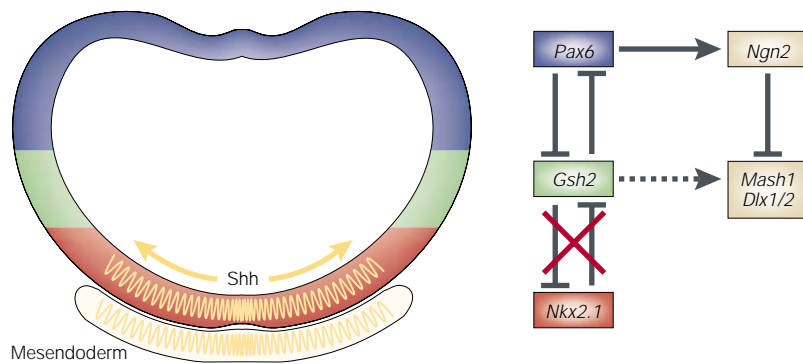


Figure 4 | Homeodomain and bHLH genetic interactions in telencephalic development. Schematic of a coronal section of the mouse telencephalon at 26 somites (about embryonic day 9.5), showing the expression pattern of the homeodomain transcription factors *Nkx2.1*, *Gsh2* and *Pax6*. Sources of both ventral telencephalic and mesodermal *Shh* are also shown in this schematic. At this age, shortly after *Gsh2* is first detected in the lateral telencephalon, expression of these genes is mostly non-overlapping. Genetic analysis has revealed that while *Pax6* and *Gsh2* act in a cross-repressive manner, *Nkx2.1* and *Gsh2* do not. Furthermore, *Pax6* may regulate the function of the basic helix–loop–helix (bHLH) transcription factor, *Ngn2*, which in turn regulates *Mash1* and *Dlx1/Dlx2* gene function.

Similarly, in the absence of *Nkx2.1*, the ventral-most aspect of the telencephalon — the MGE — becomes trans-fated to that of the adjacent, more dorsal LGE⁷⁶. However, despite the apparent conservation of the function of *Nkx/vnd* genes in the generation of the ventral-most aspect of the neuraxis, how these genes regulate, and in turn are regulated by, other genes differs between invertebrates and vertebrates. For example, in *Drosophila*, *vnd* and *ind* mutually repress the expression of each other^{71,72}. However, *Gsh2* (the *ind* homologue) and *Nkx2.1* (the *vnd* homologue), which initially maintain complementary patterns of expression, seem to have no direct effect on each other's expression (FIG. 4). Specifically, the loss of *Gsh2* (or the combined loss of *Gsh1* and *Gsh2*) does not result in an expansion of *Nkx2.1* expression, nor does the loss of *Nkx2.1* result in the precocious expression of *Gsh2* in ventral regions (J.G.C. and G.F., unpublished observations). Interestingly, however, *Gsh2* and *Pax6* (a homologue of the *Drosophila ey* gene), the complementary expression of which forms a sharp border at the intermediate–dorsal boundary (LGE–cortex), do cross-repress one another (J.G.C. and G.F., unpublished observations; FIG. 4). Loss of *Gsh2* function leads to an expansion of *Pax6* expression (as well as other dorsally restricted genes, most notably *Ngn2*) into the intermediate, LGE domain^{77–79}. Conversely, loss of *Pax6* function in small-eye (*sey*) mice leads to an expansion of *Gsh2* into the *Pax6* domain^{78,79}. In the *Drosophila* nerve cord, the dorsal repression of *ind* is known to be mediated by *msh*. However, on the basis of findings in mammals, it would also be interesting to explore the interactions between *ind* and *ey* in the establishment of the intermediate–dorsal boundary in the fly nervous system. It is worth noting that, in the vertebrate nervous system, the cross-repression of *Pax6* and *Gsh2* is specific to the telencephalon; in the spinal cord, their expression domains overlap⁷³. This, again, emphasizes

that to understand the role of the various genes in the patterning of the telencephalon, this structure must be studied directly.

Normal patterning of the telencephalon also seems to require the function of conserved members of the bHLH transcription-factor gene family. These genes include *Mash1* (a homologue of the genes in the *Drosophila achaete-scute complex*), which is expressed at its highest levels in the ventral telencephalon (MGE and LGE), and *Ngn1/Ngn2* (homologues of the *Drosophila atonal* genes), the expression of which is restricted to the dorsal telencephalon¹⁰. The absence of either *Mash1* or *Ngn1/Ngn2* gene function in mice leads to defects in DV patterning, showing the indispensable role of these bHLH genes in the normal allocation of territories in the telencephalon.

The regulatory mechanisms of bHLH and homeodomain regional gene expression are interdependent (FIG. 4). For example, in the absence of *Gsh2*, the expression of *Mash1* is lost in the LGE, and the expression of *Ngn2*, along with that of *Pax6*, is expanded into the intermediate region. In *Pax6* mutant mice, on the other hand, *Ngn2* expression is lost in the lateral cortex and *Mash1* expression expands into this region^{77–79}. Conversely, in *Mash1*-null mice, *Nkx2.1* expression is lost in the rostral MGE⁸⁰, whereas in *Ngn1/Ngn2* compound null mice, the normal ventral restriction in the expression of homeodomain genes, such as *Dlx2*, is compromised, resulting in their expansion into dorsal regions⁸¹. Collectively, these data indicate that correct DV patterning of the telencephalon is dependent not only on the interactions of conserved homeodomain genes, but also on their interactions with the bHLH genes. In addition to their role in patterning, *Mash1* and *Dlx1/Dlx2* regulate specific aspects of neurogenesis. Genetic loss-of-function studies have revealed that *Mash1* and *Dlx1/Dlx2* are required for the generation of early- and late-born subpallial progenitors, respectively^{80,82–84}. It is notable that the bHLH genes are also regulated by components of the Notch/lateral signalling pathway^{85,86}. Indeed, abnormalities in neural differentiation are hallmarks of mutants that lack either *Mash1* or *Ngn1/Ngn2* gene function. Given that an obligate part of the telencephalic patterning mechanism is the generation of regional differences in the regulation of proliferation and differentiation, it seems likely that the bHLH genes provide an important juncture by which these two processes are linked. An important area of future research will be to delineate more clearly the connection between the control of regional patterning and the means by which proliferation in dorsal versus ventral regions of the telencephalon is coordinately regulated.

One of the specific functions of *Shh* is to induce oligodendrocyte development in ventral regions of the central nervous system, including the telencephalon^{87,88}. Two novel bHLH factors, *Olig1* and *Olig2*, have been identified that are essential for the specification of oligodendrocytes in response to *Shh* signalling^{89,90}. Recently, single and compound null alleles of these genes have been generated^{91,92}. These studies indicate that oligodendrocytes and motor neurons in the spinal

cord have an absolute requirement for these genes, and that they act in combination with *Nkx2.2* and *Ngn1*, respectively, to establish these populations^{93–95}. *Olig2* is expressed widely in the ventral telencephalon, but the phenotype that results from the loss of this gene in this region has not yet been reported in detail. Given that both oligodendrocytes and interneurons originate from ventral telencephalic regions^{87,96}, and might share a common precursor⁹⁷, it will be of interest to see how the loss of *Olig1/Olig2* affects the generation of these cell types. More broadly, these genes no doubt provide a good example of the kind of combinatorial code that is required for the generation of specific cell types; for example, in spinal cord, progenitors that co-express *Olig1/Olig2* and *Nkx2.1* become oligodendrocytes, whereas progenitors that co-express *Olig1/Olig2* and *Ngn1* become motor neurons^{93–95}. Indeed, an important aim, which further analysis of the telencephalon must address, is to uncover the logic of how combinatorial codes of homeodomain and bHLH genes act to establish specific neural populations within the telencephalon.

Tangential migration broadens regional diversity By E12.5 in mice, as a result of regional patterning, the telencephalon develops a series of characteristic proliferative zones. In the dorsal telencephalon, both cortical and hippocampal regions become evident, and three distinct eminences appear ventrally: the MGE, the LGE anteriorly, and the caudal ganglionic eminence (CGE) posteriorly^{98–100}. Until recently, these different regions of the forebrain were thought to develop as independent compartments, with cortical cells originating entirely from the cortical ventricular zone, and the striatum, globus pallidus and amygdala arising from the ventral eminences^{101–103}. A wealth of genetic, lineage-tracing and fate-mapping analyses has revealed the situation to be considerably more complicated^{102,103}. Although many details remain to be worked out, it now seems clear that extensive mixing of progeny occurs through tangential migration, with cells from the MGE and CGE ultimately populating the cortex, and cortically derived neurons migrating ventrally to invade the amygdala. The logic of why such a baroque scheme of development has been selected for probably reflects the fact that different progenitor zones generate specific subsets of neural cell types. For example, the MGE and CGE make large populations of interneurons and oligodendrocytes, whereas the LGE, hippocampal and cortical proliferative zones generate primarily projection neurons. So, regional patterning not only generates specific telencephalic structures, but also acts as a means of producing large populations of particular neuronal subtypes.

Summary and future directions

Work over the past ten years has begun to give us a mechanistic understanding of telencephalic development. In summary, telencephalic development follows a discrete series of steps. First, around the time of gastrulation, an interplay between posteriorizing

signals (such as FGFs, Wnts and retinoids) and anteriorizing factors (such as cerberus and dickkopf) results in the polarization of the nervous system^{12,14}. Anterior neural tissue later becomes further subdivided into the diencephalon and telencephalon, at least partially by the graded modulation of Wnt signalling^{31–33}. This is followed by the emergence of regional patterning in the telencephalon itself, which requires the action of Shh, as well as other extrinsic factors, possibly including FGF, BMPs, Wnts, retinoids and Nodal signalling. Although there might be some redundancy between the factors that establish positional information within the telencephalon (as shown by the persistence of DV patterning in the telencephalon of *Shh/Gli3* compound mutants⁹), such signalling seems to converge on the induction of a combinatorial code of homeodomain and bHLH transcription factors within progenitors at different DV positions¹⁰. Notably, this code results not only in the emergence of regional territories, but also in characteristic patterns of proliferation in dorsal versus ventral regions of the telencephalon. In addition, it seems that specific populations of cell types, including interneurons, oligodendrocytes and projection neurons, are each born in different telencephalic regions and become distributed appropriately only later in development, through characteristic patterns of tangential migration.

Obviously, there are many unanswered questions that relate to each of these steps. For example, we have not yet characterized the full range of extrinsic factors that act to establish the telencephalon and its subdivisions. Similarly, it seems certain that further transcription factors that contribute to the combinatorial code, as well as the logic by which this code results in the generation of specific subpopulations, remain to be discovered. Furthermore, recent fate-mapping efforts indicate that tangential migration within the telencephalon is widespread and includes dorsal-to-ventral migration of neurons, in addition to the well-characterized ventral-to-dorsal migration of interneurons. Nonetheless, it is clear that the signalling cascades for establishing telencephalic pattern are widely used in other regions of the embryo, as well as across species. Therefore, the molecular requirements for generating brain structures are variations of mechanisms used elsewhere. The specificity that allows the development of the telencephalon involves a unique combination and ordering of these common signals in time and space. Now that a broad outline of telencephalic development is crystallizing, it seems likely that further resolution of the underlying mechanisms will rely on a direct examination of this region, using the growing complement of cellular and genetic approaches that are available.

Note added in proof

A recent paper by Lupo *et al.*¹⁰⁴ has implicated the inhibition of Wnts and BMPs by cerberus as a crucial step towards the generation of dorsal telencephalic identity.

1. Younossi-Hartenstein, A. *et al.* Control of early neurogenesis of the *Drosophila* brain by the head gap genes *Ill*, *otd*, *ems*, and *bitd*. *Dev. Biol.* **182**, 270–283 (1997).
2. Reichert, H. & Simeone, A. Conserved usage of gap and homeotic genes in patterning the CNS. *Curr. Opin. Neurobiol.* **9**, 589–595 (1999).
3. Tanabe, Y. & Jessell, T. M. Diversity and pattern in the developing spinal cord. *Science* **274**, 1115–1123 (1996).
4. Edlund, T. & Jessell, T. M. Progression from extrinsic to intrinsic signaling in cell fate specification: a view from the nervous system. *Cell* **96**, 211–224 (1999).
Although this is a review, it articulates the logic by which extrinsic inductive cues, such as Shh, through their induction of sets of transcription factors, are translated into intrinsic cell identities.
5. Ericsson, J. *et al.* Sonic hedgehog induces the differentiation of ventral forebrain neurons: a common signal for ventral patterning within the neural tube. *Cell* **81**, 747–756 (1995).
6. Chiang, C. *et al.* Cyclopia and defective axial patterning in mice lacking Sonic hedgehog gene function. *Nature* **383**, 407–413 (1996).
This was the first characterization of a Shh-null allele, showing the requirement for Shh signalling throughout the central nervous system, including the telencephalon.
7. Kohtz, J. D., Baker, D. P., Corte, G. & Fishell, G. Regionalization within the mammalian telencephalon is mediated by changes in responsiveness to Sonic Hedgehog. *Development* **125**, 5079–5089 (1998).
This paper and reference 39 show that the Shh effects in the telencephalon are dependent on timing rather than on Shh concentration.
8. Gaiano, N., Kohtz, J. D., Turnbull, D. H. & Fishell, G. A method for rapid gain-of-function studies in the mouse embryonic nervous system. *Nature Neurosci.* **2**, 812–819 (1999).
9. Rallu, M. *et al.* Dorsal-ventral patterning is established in the telencephalon of mutants lacking both Gli3 and Hedgehog signaling. *Development* **129**, 4963–4974 (2002).
This paper shows that in the absence of both Shh (or Smo) and Gli3 gene activity, general aspects of DV patterning in the telencephalon are established, indicating that a Shh-independent pathway acts in the establishment of DV patterning in the telencephalon.
10. Schuurmans, C. & Guillemot, F. Molecular mechanisms underlying cell fate specification in the developing telencephalon. *Curr. Opin. Neurobiol.* **12**, 26–34 (2002).
11. de Souza, F. S. & Niehrs, C. Anterior endoderm and head induction in early vertebrate embryos. *Cell Tissue Res.* **300**, 207–217 (2000).
12. Stern, C. D. Initial patterning of the central nervous system: how many organizers? *Nature Rev. Neurosci.* **2**, 92–98 (2001).
13. Beddington, R. S. & Robertson, E. J. Anterior patterning in mouse. *Trends Genet.* **14**, 277–284 (1998).
14. Lu, C. C., Brennan, J. & Robertson, E. J. From fertilization to gastrulation: axis formation in the mouse embryo. *Curr. Opin. Genet. Dev.* **11**, 384–392 (2001).
15. Thomas, P. & Beddington, R. Anterior primitive endoderm may be responsible for patterning the anterior neural plate in the mouse embryo. *Curr. Biol.* **6**, 1487–1496 (1996).
This paper shows that activity in the anterior visceral ectoderm is required for anterior neural patterning in the mouse, showing that, in mammals, activities beyond those found in the organizer are required for the establishment/maintenance of anterior neural character.
16. Beddington, R. S. & Robertson, E. J. Axis development and early asymmetry in mammals. *Cell* **96**, 195–209 (1999).
17. Knoetgen, H., Teichmann, U. & Kessel, M. Head-organizing activities of endodermal tissues in vertebrates. *Cell. Mol. Biol.* **45**, 481–492 (1999).
18. Nieuwkoop, P. D. & Nigtevecht, G. V. Neural activation and transformation in explants of competent ectoderm under the influence of fragments of anterior notochord in Urodeles. *J. Embryol. Exp. Morph.* **2**, 175–193 (1954).
19. Sasai, Y. & De Robertis, E. M. Ectodermal patterning in vertebrate embryos. *Dev. Biol.* **182**, 5–20 (1997).
20. Moon, R. T. & Kimelman, D. From cortical rotation to organizer gene expression: toward a molecular explanation of axis specification in *Xenopus*. *Bioessays* **20**, 536–545 (1998).
21. Altmann, C. R. & Brivanlou, A. H. Neural patterning in the vertebrate embryo. *Int. Rev. Cytol.* **203**, 447–482 (2001).
22. Schier, A. F. Axis formation and patterning in zebrafish. *Curr. Opin. Genet. Dev.* **11**, 393–404 (2001).
23. Bouwmeester, T., Kim, S., Sasai, Y., Lu, B. & De Robertis, E. M. Cerberus is a head-inducing secreted factor expressed in the anterior endoderm of Spemann's organizer. *Nature* **382**, 595–601 (1996).
24. Glinka, A. *et al.* Dickkopf-1 is a member of a new family of secreted proteins and functions in head induction. *Nature* **391**, 357–362 (1998).
25. Schier, A. F. & Shen, M. M. Nodal signalling in vertebrate development. *Nature* **403**, 385–389 (2000).
26. Thomsen, G. H. Antagonism within and around the organizer: BMP inhibitors in vertebrate body patterning. *Trends Genet.* **13**, 209–211 (1997).
27. Harland, R. Neural induction. *Curr. Opin. Genet. Dev.* **10**, 357–362 (2000).
28. Wessely, O. & De Robertis, E. M. Neural plate patterning by secreted signals. *Neuron* **33**, 489–491 (2002).
29. Shimamura, K. & Rubenstein, J. L. Inductive interactions direct early regionalization of the mouse forebrain. *Development* **124**, 2709–2718 (1997).
30. Houart, C., Westerfield, M. & Wilson, S. W. A small population of anterior cells patterns the forebrain during zebrafish gastrulation. *Nature* **391**, 788–792 (1998).
This work shows that non-neural cells anterior to the neural plate can induce anterior tissue (ANB cells) to adopt a telencephalic phenotype (see also reference 33).
31. Heisenberg, C. P. *et al.* A mutation in the Gsk3-binding domain of zebrafish Masterblind/Axin1 leads to a fate transformation of telencephalon and eyes to diencephalon. *Genes Dev.* **15**, 1427–1434 (2001).
32. Masai, I. *et al.* *floating head* and *masterblind* regulate neuronal patterning in the roof of the forebrain. *Neuron* **18**, 43–57 (1997).
33. Houart, C. *et al.* Establishment of the telencephalon during gastrulation by local antagonism of Wnt signaling. *Neuron* **35**, 255–265 (2002).
This paper revealed that Tlc, a Frizzled-related Wnt antagonist, is present in ANB cells, and is necessary and sufficient for the establishment of forebrain identities.
34. Briscoe, J. & Ericson, J. The specification of neuronal identity by graded Sonic Hedgehog signalling. *Semin. Cell Dev. Biol.* **10**, 353–362 (1999).
35. Ingham, P. W. & McMahon, A. P. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev.* **15**, 3059–3087 (2001).
36. Pabst, O., Herbrand, H., Takuma, N. & Arnold, H. H. NKX2 gene expression in neuroectoderm but not in mesodermally derived structures depends on sonic hedgehog in mouse embryos. *Dev. Genes Evol.* **210**, 47–50 (2000).
37. Echelard, Y. *et al.* Sonic hedgehog, a member of a family of putative signaling molecules, is implicated in the regulation of CNS polarity. *Cell* **75**, 1417–1430 (1993).
38. Shimamura, K., Hartigan, D. J., Martinez, S., Puelles, L. & Rubenstein, J. L. Longitudinal organization of the anterior neural plate and neural tube. *Development* **121**, 3923–3933 (1995).
39. Gunhaga, L., Jessell, T. M. & Edlund, T. Sonic hedgehog signaling at gastrula stages specifies ventral telencephalic cells in the chick embryo. *Development* **127**, 3283–3293 (2000).
40. Aza-Blanc, P. & Kornberg, T. B. Ci: a complex transducer of the hedgehog signal. *Trends Genet.* **15**, 458–462 (1999).
41. Matisse, M. P. & Joyner, A. L. Gli genes in development and cancer. *Oncogene* **18**, 7852–7859 (1999).
42. Park, H. L. *et al.* Mouse *Gli3* mutants are viable but have defects in SHH signaling in combination with a *Gli2* mutation. *Development* **127**, 1593–1605 (2000).
43. Theil, T., Alvarez-Bolado, G., Walter, A. & Ruther, U. *Gli3* is required for *Emx* gene expression during dorsal telencephalon development. *Development* **126**, 3561–3571 (1999).
44. Grove, E. A., Tole, S., Limon, J., Yip, L. & Ragsdale, C. W. The hem of the embryonic cerebral cortex is defined by the expression of multiple *Wnt* genes and is compromised in *Gli3*-deficient mice. *Development* **125**, 2315–2325 (1998).
45. Litingtung, Y. & Chiang, C. Specification of ventral neuron types is mediated by an antagonistic interaction between Shh and Gli3. *Nature Neurosci.* **3**, 979–985 (2000).
This study presents the first analysis of Shh/Gli3 double mutants, showing that many aspects of the spinal cord phenotype in Shh mutants can be rescued by combined removal of Shh and Gli3.
46. Wang, B., Fallon, J. F. & Beachy, P. A. Hedgehog-regulated processing of Gli3 produces an anterior/posterior repressor gradient in the developing vertebrate limb. *Cell* **118**, 423–434 (2000).
47. Litingtung, Y., Dahn, R. D., Li, Y., Fallon, J. F. & Chiang, C. *Shh* and *Gli3* are dispensable for limb skeleton formation but regulate digit number and identity. *Nature* **418**, 979–983 (2002).
48. Bai, C. B. & Joyner, A. L. *Gli1* can rescue the *in vivo* function of *Gli2*. *Development* **128**, 5161–5172 (2001).
49. Liem, K. F. Jr, Tremml, G., Roelink, H. & Jessell, T. M. Dorsal differentiation of neural plate cells induced by BMP-mediated signals from epidermal ectoderm. *Cell* **82**, 969–979 (1995).
50. Liem, K. F. Jr, Jessell, T. M. & Briscoe, J. Regulation of the neural patterning activity of sonic hedgehog by secreted BMP inhibitors expressed by notochord and somites. *Development* **127**, 4855–4866 (2000).
51. Timmer, J. R., Wang, C. & Niswander, L. BMP signalling patterns the dorsal and intermediate neural tube via regulation of homeobox and helix-loop-helix transcription factors. *Development* **129**, 2459–2472 (2002).
52. Tole, S., Ragsdale, C. W. & Grove, E. A. Dorsal-ventral patterning of the telencephalon is disrupted in the mouse mutant extra-toes. *J. Biol.* **217**, 254–265 (2000).
53. Monuki, E. S., Porter, F. D. & Walsh, C. A. Patterning of the dorsal telencephalon and cerebral cortex by a roof plate-Lhx2 pathway. *Neuron* **32**, 591–604 (2001).
54. Ohkubo, Y., Chiang, C. & Rubenstein, J. L. Coordinate regulation and synergistic actions of BMP4, SHH and FGF8 in the rostral prosencephalon regulate morphogenesis of the telencephalic and optic vesicles. *Neuroscience* **111**, 1–17 (2002).
55. Theil, T., Aydin, S., Koch, S., Grotewold, L. & Ruther, U. Wnt and Bmp signalling cooperatively regulate graded *Emx2* expression in the dorsal telencephalon. *Development* **129**, 3045–3054 (2002).
56. Hébert, J. M., Mishina, Y. & McConnell, S. K. BMP signaling is required locally to pattern the dorsal telencephalic midline. *Neuron* **35**, 1029–1041 (2002).
57. Dale, J. K. *et al.* Cooperation of BMP7 and SHH in the induction of forebrain ventral midline cells by prechordal mesoderm. *Cell* **90**, 257–269 (1997).
58. Shanmugalingam, S. *et al.* *Ace/Fgf8* is required for forebrain commissure formation and patterning of the telencephalon. *Development* **127**, 2549–2561 (2000).
59. Galceran, J., Miyashita-Lin, E. M., Devaney, E., Rubenstein, J. L. & Grosschedl, R. Hippocampus development and generation of dentate gyrus granule cells is regulated by Lef1. *Development* **127**, 469–482 (2000).
60. Lee, S. M., Tole, S., Grove, E. & McMahon, A. P. A local Wnt-3a signal is required for development of the mammalian hippocampus. *Development* **127**, 457–467 (2000).
61. Pierani, A., Brenner-Morton, S., Chiang, C. & Jessell, T. M. A sonic hedgehog-independent, retinoid-activated pathway of neurogenesis in the ventral spinal cord. *Cell* **97**, 903–915 (1999).
62. Toresson, H., Mata de Urquiza, A., Fagerstrom, C., Perlmann, T. & Campbell, K. Retinoids are produced by glia in the lateral ganglionic eminence and regulate striatal neuron differentiation. *Development* **126**, 1317–1326 (1999).
63. Li, H. *et al.* A retinoic acid synthesizing enzyme in ventral retina and telencephalon of the embryonic mouse. *Mech. Dev.* **95**, 283–289 (2000).
64. LaMantia, A. S., Colbert, M. C. & Linney, E. Retinoic acid induction and regional differentiation prefigure olfactory pathway formation in the mammalian forebrain. *Neuron* **10**, 1035–1048 (1993).
65. Shinya, M., Koshida, S., Sawada, A., Kuroiwa, A. & Takeda, H. Fgf signalling through MAPK cascade is required for development of the subpallial telencephalon in zebrafish embryos. *Development* **128**, 4153–4164 (2001).
66. Rohr, K. B., Barth, K. A., Varga, Z. M. & Wilson, S. W. The nodal pathway acts upstream of hedgehog signaling to specify ventral telencephalic identity. *Neuron* **29**, 341–351 (2001).
67. Cornell, R. A. & Ohlen, T. V. *vnd/nkx*, *ind/gsh*, and *msh/msx*: conserved regulators of dorsoventral neural patterning? *Curr. Opin. Neurobiol.* **10**, 63–71 (2000).
68. Bertrand, N., Castro, D. S. & Guillemot, F. Proneural genes and the specification of neural cell types. *Nature Rev. Neurosci.* **3**, 517–530 (2002).
69. Jessell, T. M. & Sanes, J. R. Development. The decade of the developing brain. *Curr. Opin. Neurobiol.* **10**, 599–611 (2000).
70. Lu, S., Bogarad, L. D., Murtha, M. T. & Ruddle, F. H. Expression pattern of a murine homeobox gene, *Dbx*, displays extreme spatial restriction in embryonic forebrain and spinal cord. *Proc. Natl. Acad. Sci. USA* **89**, 8053–8057 (1992).
71. Chu, H., Parras, C., White, K. & Jimenez, F. Formation and specification of ventral neuroblasts is controlled by *vnd* in *Drosophila* neurogenesis. *Genes Dev.* **12**, 3613–3624 (1998).
72. McDonald, J. A. Dorsal-ventral patterning in the *Drosophila* central nervous system: the *vnd* homeobox gene specifies ventral column identity. *Genes Dev.* **12**, 3603–3612 (1998).
This work, together with references 69 and 71, shows that there is conservation in the genes that establish ventral identity in the *Drosophila* and vertebrate central nervous system.

73. Weiss, J. B. Dorsal-ventral patterning in the *Drosophila* central nervous system: the *intermediate neuroblasts defective* homeobox gene specifies intermediate column identity. *Genes Dev.* **12**, 3591–3602 (1998).
74. Marquardt, T. & Gruss, P. Generating neuronal diversity in the retina: one for nearly all. *Trends Neurosci.* **25**, 32–38 (2002).
75. Briscoe, J. Homeobox gene *Nkx2.2* and specification of neuronal identity by graded Sonic hedgehog signalling. *Nature* **398**, 622–627 (1999).
76. Sussel, L., Marin, O., Kimura, S. & Rubenstein, J. L. Loss of *Nkx2.7* homeobox gene function results in a ventral to dorsal molecular respecification within the basal telencephalon: evidence for a transformation of the pallidum into the striatum. *Development* **126**, 3359–3370 (1999). **This paper reports the remarkable finding that MGE (ventral-most telencephalic fate) is transformed into LGE (lateral telencephalic fate) in the absence of *Nkx2.1* gene function.**
77. Corbin, J. G., Gaiano, N., Machold, R. P., Langston, A. & Fishell, G. The *Gsh2* homeodomain gene controls multiple aspects of telencephalic development. *Development* **127**, 5007–5020 (2000).
78. Toresson, H., Potter, S. S. & Campbell, K. Genetic control of dorsal-ventral identity in the telencephalon: opposing roles for *Pax6* and *Gsh2*. *Development* **127**, 4361–4371 (2000).
79. Yun, K., Potter, S. & Rubenstein, J. L. *Gsh2* and *Pax6* play complementary roles in dorsoventral patterning of the mammalian telencephalon. *Development* **128**, 193–205 (2001).
80. Casarosa, S., Fode, C. & Guillemot, F. *Mash1* regulates neurogenesis in the ventral telencephalon. *Development* **126**, 525–534 (1999).
81. Fode, C. *et al.* A role for neural determination genes in specifying the dorsoventral identity of telencephalic neurons. *Genes Dev.* **14**, 67–80 (2000).
82. Horton, S., Meredith, A., Richardson, J. A. & Johnson, J. E. Correct coordination of neuronal differentiation events in ventral forebrain requires the bHLH factor MASH1. *Mol. Cell Neurosci.* **14**, 355–369 (1999).
83. Anderson, S. A. *et al.* Mutations of the homeobox genes *Dlx-1* and *Dlx-2* disrupt the striatal subventricular zone and differentiation of late born striatal neurons. *Neuron* **19**, 27–37 (1997).
84. Yun, K. *et al.* Modulation of the notch signaling by *Mash1* and *Dlx1/2* regulates sequential specification and differentiation of progenitor cell types in the subcortical telencephalon. *Development* **129**, 5029–5040 (2002).
85. Ma, Q., Kintner, C. & Anderson, D. J. Identification of neurogenin, a vertebrate neuronal determination gene. *Cell* **87**, 43–52 (1996).
86. Ma, Q., Sommer, L., Cserjesi, P. & Anderson, D. J. *Mash1* and *neurogenin1* expression patterns define complementary domains of neuroepithelium in the developing CNS and are correlated with regions expressing notch ligands. *J. Neurosci.* **17**, 3644–3652 (1997).
87. Nery, S., Wichterle, H. & Fishell, G. Sonic hedgehog contributes to oligodendrocyte specification in the mammalian forebrain. *Development* **128**, 527–540 (2001). **This paper and reference 82 show that Shh is involved in the generation of oligodendrocytes in the telencephalon, and that, like interneurons, many cortical oligodendrocytes probably originate in the ventral telencephalon.**
88. Tekki-Kessaris, N. Hedgehog-dependent oligodendrocyte lineage specification in the telencephalon. *Development* **128**, 2545–2554 (2001).
89. Lu, Q. R. Sonic hedgehog-regulated oligodendrocyte lineage genes encoding bHLH proteins in the mammalian central nervous system. *Neuron* **25**, 317–329 (2000).
90. Zhou, Q., Wang, S. & Anderson, D. J. Identification of a novel family of oligodendrocyte lineage-specific basic helix-loop-helix transcription factors. *Neuron* **25**, 331–343 (2000).
91. Lu, Q. R. Common developmental requirement for *Olig* function indicates a motor neuron/oligodendrocyte connection. *Cell* **109**, 75–86 (2002).
92. Zhou, Q. & Anderson, D. J. The bHLH transcription factors OLIG2 and OLIG1 couple neuronal and glial subtype specification. *Cell* **109**, 61–73 (2002).
93. Novitsch, B. G., Chen, A. I. & Jessell, T. M. Coordinate regulation of motor neuron subtype identity and pan-neuronal properties by the bHLH repressor *Olig2*. *Neuron* **31**, 773–789 (2001).
94. Sun, T. *Olig* bHLH proteins interact with homeodomain proteins to regulate cell fate acquisition in progenitors of the ventral neural tube. *Curr. Biol.* **11**, 1413–1420 (2001).
95. Zhou, Q., Choi, G. & Anderson, D. J. The bHLH transcription factor *Olig2* promotes oligodendrocyte differentiation in collaboration with *Nkx2.2*. *Neuron* **31**, 791–807 (2001).
96. Anderson, S. A., Eisenstat, D. D., Shi, L. & Rubenstein, J. L. Interneuron migration from basal forebrain to neocortex: dependence on *Dlx* genes. *Science* **278**, 474–476 (1997). **Provided the proof that *Dlx*-expressing cortical interneurons originate in ventral regions of the telencephalon.**
97. He, W., Ingraham, C., Rising, L., Goderie, S. & Temple, S. Multipotent stem cells from the mouse basal forebrain contribute GABAergic neurons and oligodendrocytes to the cerebral cortex during embryogenesis. *J. Neurosci.* **21**, 8854–8862 (2001).
98. Fishell, G. Regionalization in the mammalian telencephalon. *Curr. Opin. Neurobiol.* **7**, 62–69 (1997).
99. Wilson, S. O. W. & Rubenstein, J. L. Induction and dorsoventral patterning of the telencephalon. *Neuron* **28**, 641–651 (2000).
100. Nery, S., Fishell, G. & Corbin, J. G. The caudal ganglionic eminence is a novel source of distinct cortical and subcortical cell populations. *Nature Neurosci.* (in the press).
101. Parnavelas, J. G. The origin and migration of cortical neurones: new vistas. *Trends Neurosci.* **23**, 126–131 (2000).
102. Corbin, J. G., Nery, S. & Fishell, G. Telencephalic cells take a tangent: non-radial migration in the mammalian forebrain. *Nature Neurosci.* **4**, 1177–1182 (2001).
103. Marin, O. & Rubenstein, J. L. A long, remarkable journey: tangential migration in the telencephalon. *Nature Rev. Neurosci.* **2**, 780–790 (2001).
104. Lupo, G., Harris, W. A., Bar sacchi, G. & Vignali, A. Induction and patterning of the telencephalon in *Xenopus laevis*. *Development* **129**, 5421–5436 (2002).

Acknowledgements

We thank all members of the Fishell lab for their critical reading of this review. We are also grateful to S. Wilson for his many helpful suggestions and for clarification of our worst misstatements. Our work is supported by the National Institutes of Health (NIH), a March of Dimes basic research grant and a Children's Brain Tumor Foundation grant to G.F.; and by postdoctoral grants from l'Association pour la Recherche Contre le Cancer to M.R., from the American Cancer Society to N.G. and from the NIH to J.C. and R.M. Finally, including all the important findings on the telencephalon in a single review is an impossible task. Where possible, we have cited reviews, rather than primary sources, to be as inclusive as possible. We hope that the many authors of important papers on the subject that we have failed to mention will forgive us for our oversights.

Online links

DATABASES

The following terms in this article are linked online to:

Entrez: <http://ncbi.nlm.nih.gov/Entrez/>
 cerberus | dickkopf | Tlc | Wnt
FlyBase: <http://flybase.bio.indiana.edu/>
 achaete-scute complex | atonal | cubitus interruptus | eyeless | H2.0 | Hedgehog | ind | msh | Smoothed | vnd
LocusLink: <http://www.ncbi.nlm.nih.gov/LocusLink/>
 Axin | Bf1 | *Bmp2b* | BMP4 | BMP7 | chordin | *Dbx1* | *Dbx2* | *Dlx1* | *Dlx2* | Emx1 | FGF3 | FGF8 | follistatin | Frzb | *Gli1* | *Gli2* | *Gli3* | *Gsh1* | *Gsh2* | *Hesx1* | Ihh | *Lim1* | *Mash1* | *Ngn1* | *Ngn2* | *Nkx2.1* | *Nkx2.2* | *Nkx6.1* | Nodal | noggin | Notch | *Olig1* | *Olig2* | *Otx2* | *Pax6* | Shh | Smo | TGF- β

FURTHER INFORMATION

Encyclopedia of Life Sciences: <http://www.els.net/>
 bone morphogenetic proteins and their receptors | hedgehog signalling | mammalian embryo: Wnt signalling | neural development: bHLH genes | neural subtype identity regulation | signal transduction pathways in development: Wnts and their receptors | vertebrate central nervous system: pattern formation
The Fishell Lab: <http://skirball.med.nyu.edu/groups/FishellLab/>
 Access to this interactive links box is free online.